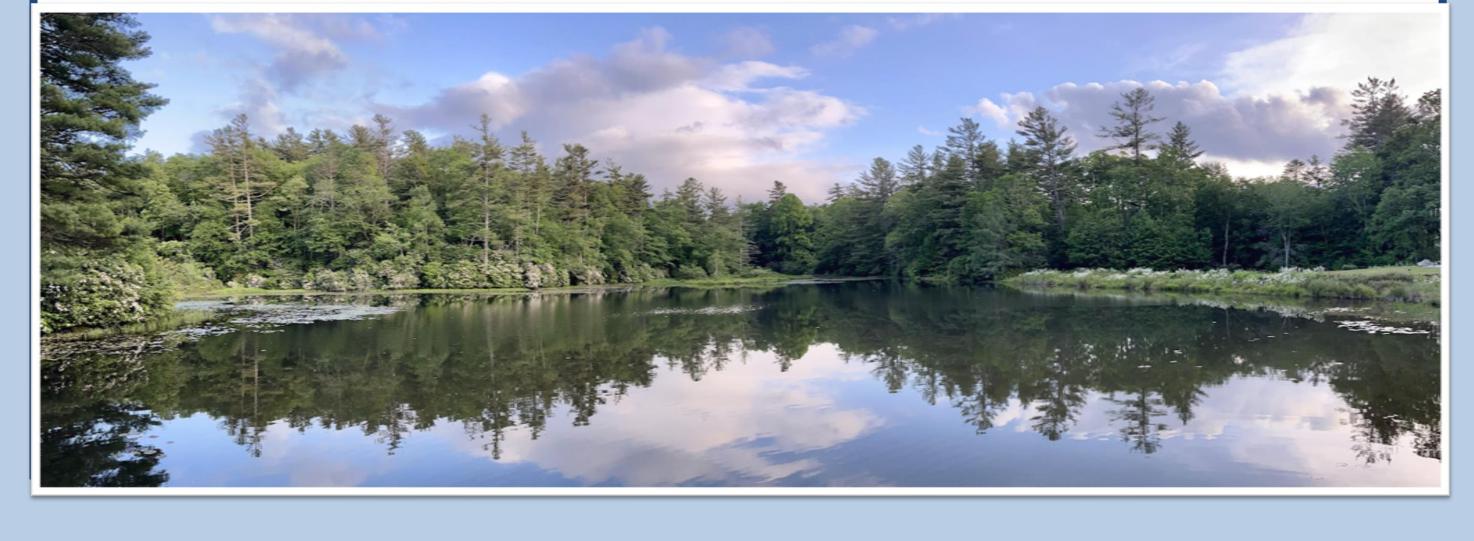


Introduction

The durable, easy to produce, and cheap nature of plastics has rendered them a widespread pollutant in the environment (Jambeck et al. 2015). Only 9% of plastics ever produced have been recycled, with 79% ending up in landfills and 12% being incinerated (Geyer et al. 2017). As these plastics degrade, they form microplastics (MPs), which are defined as particles between 1 µm and 5 mm in length (Mani et al. 2015, Valsesia et al. 2021). The ubiquity of MPs in the environment presents numerous health concerns for both ecosystems and humans. When exposed to MPs, aquatic biota experience a range of toxic molecular and physiological impacts, including alterations to immune, metabolic, growth, reproductive functions (Anbumani and Kakkar 2018, Franzellitti et al. 2019).

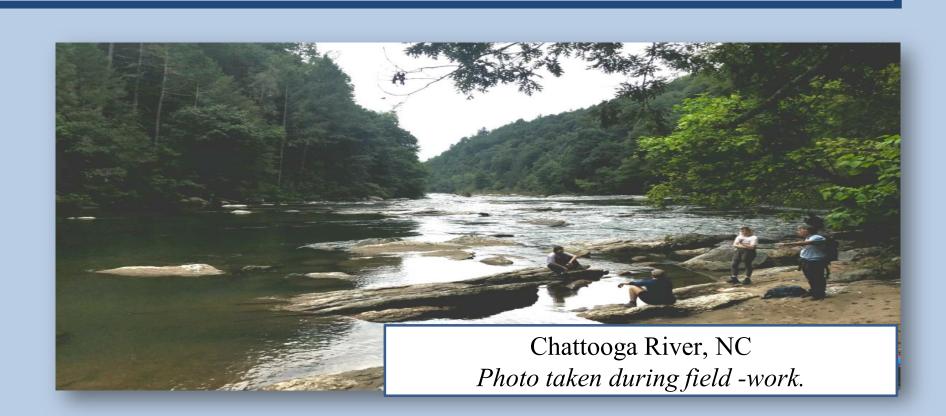
This study aims to contribute to the growing body of evidence of MP accumulation and contamination in North American freshwater systems, considering that less than 4% of MP research is focused on freshwaters (Lambert and Wagner 2018). Research of this nature is novel considering that no freshwater MP analysis in the southeastern US has been published and there is only one published study that has examined MP concentrations during storm events (Hitchcock 2020). Microplastic research regarding aquatic systems is not only vital information for the management of freshwater systems but is also vital information for those working to preserve the southern Appalachians, a well-known biodiversity hotspot (Van Sickle 1999, Simon et al. 2005).



Study Objectives

- The present study aims to determine the levels of microplastics in headwater streams in the southern Appalachians by investigating how storm events influence microplastic concentrations in streams, atmospheric deposition, and runoff, thus allowing conclusions to be made on the potential sources of contamination.
- Additional goals of this study are to **quantify and characterize** any detected microplastics to help increase the current knowledge database for the novel field of microplastic research in freshwater systems.
- Establish novel MP sampling, quality control, and characterization techniques using Raman spectroscopy.





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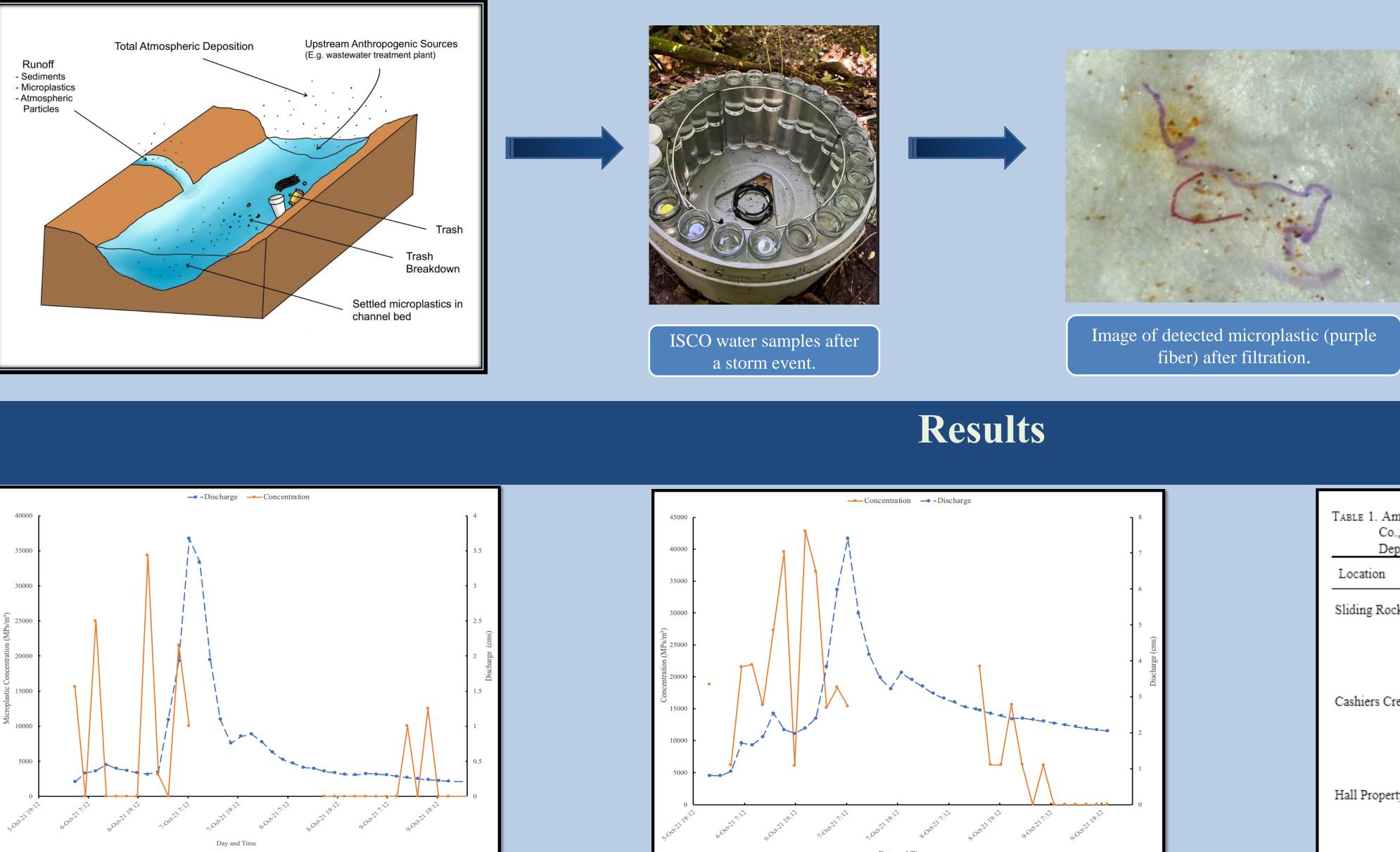
BIG RIVERS, LITTLE PLASTICS: Microplastic concentrations and dynamics in headwater streams in the southern Appalachian mountains Chloe Hall, Jason Love (HBS), Dr. Jerry Miller (WCU), & Noa Meiri (UNC) UNC SURF Advisor: Dr. Radmila Petric

• Sampling was conducted in the Chattooga River watershed in Highlands, NC at two headwater streams above and below the local wastewater treatment plant and at designated atmospheric deposition sites (Fig. 1). Topographic and hydrologic surveying at the two stream sites was conducted regularly to calculate discharge (cms) and create representative rating curves.

When storm events with precipitation >2.5 cm of precipitation were identified in the study region, we deployed the pre-installed Teledyne ISCO full-size portable sampler (ISCO) to collect water samples at programmed intervals during base and storm flow. Weekly quality control samples using DI water were collected from the ISCOS as well as control grab samples from the stream sites.

Atmospheric deposition collection buckets were deployed in the terrain surrounding the two stream sites and were opened up for collection during storm events. All samples (stream, atmospheric, and control) were filtered using a vacuum system and then analyzed under a microscope for MP presence.

composition of the plastic.



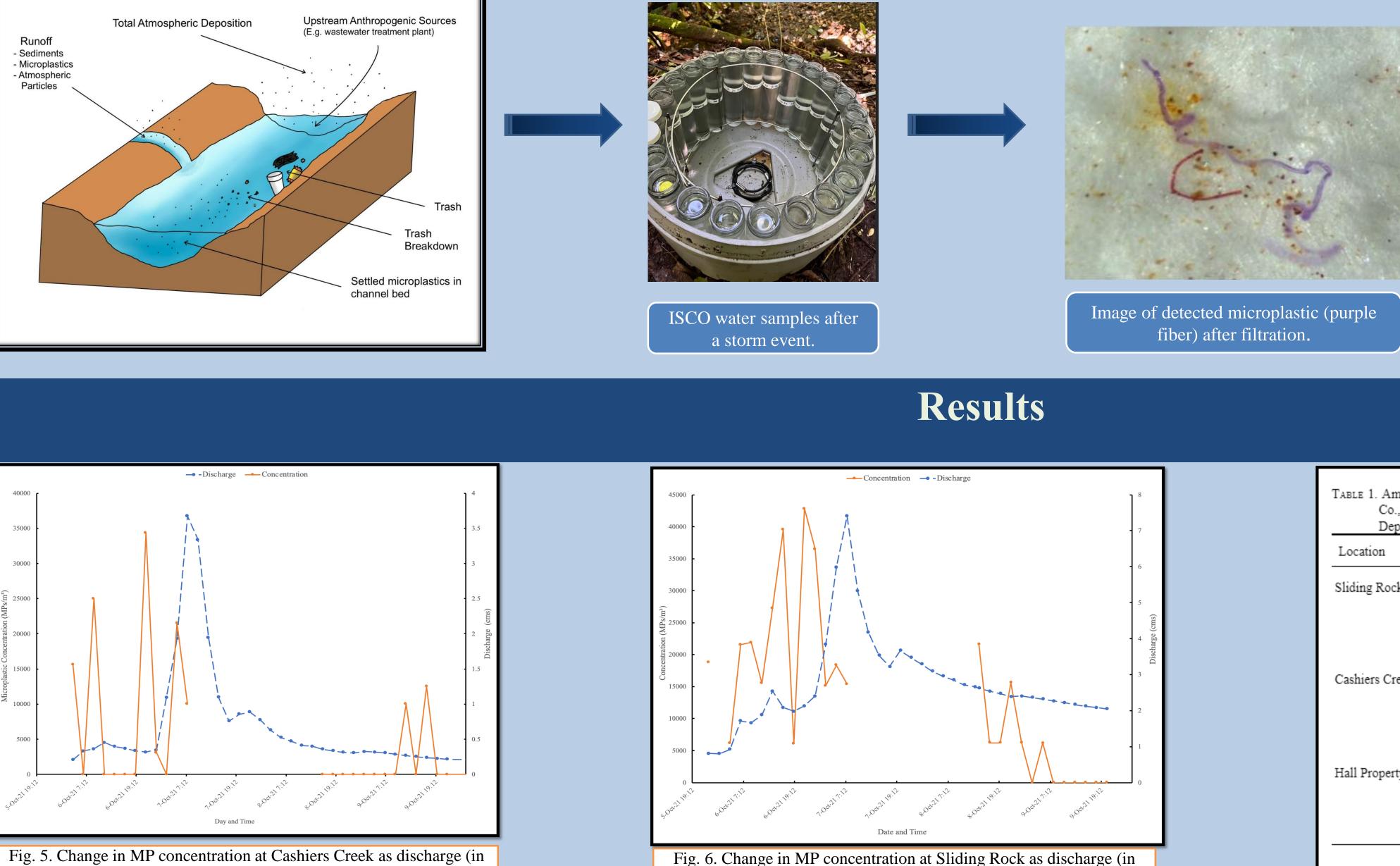


Fig. 5. Change in MP concentration at Cashiers Creek as discharge (in cubic meters per second) changed during the storm event.

deposition of microplastics into freshwater systems.

Anbumani, S., and P. Kakkar. 2018. Ecotoxicological effects of microplastics on biota: A review. Environmental Science and Pollution Research 25:14373-14396. Franzellitti, S., L. Canesi, M. Auguste, R. H. Wathsala, and E. Fabbri. 2019. Microplastic exposure and effects in aquatic organisms: A physiological perspective. Environmental Toxicology and Pharmacology 68:37-51. Geyer, R., J. R. Jambeck, and K. L. Law. 2017. Production, use, and fate of all plastics ever made. Science Advances 3:e1700782. Jambeck, J. R., Geyer, R., Wilcox, C., Siegler, T. R., Perryman, M., Andrady, A., Narayan, R., and Law, K. L. 2015. Plastic waste inputs from land into the ocean. Science 347:768–771. Lambert, S., and M. Wagner. 2018. Microplastics are contaminants of emerging concern in freshwater environments: An overview. Pages 1-23 in M. Wagner and S. Lambert, editors. Freshwater Microplastics Emerging Environmental Contaminants? Springer Nature, Cham, Switzerland. Mani, T., A. Hauk, U. Walter, and P. Burkhardt-Holm. 2015. Microplastics profile along the Rhine River. Scientific Reports 5:1-7. Simon, S. A., T. K. Collins, G. L. Kauffman, W. H. McNab, and C. J. Ulrey. 2005. Ecological zones in the southern Appalachians: First approximation. US Forest Service. Valsesia, A., J. Parot, J. Ponti, D. Mehn, R. Marino, D. Melillo, S. Muramoto, M. Verkouteren, V. A. Hackley, and P. Colpo. 2021. Detection, counting and characterization of nanoplastics in marine bioindicators: A proof of principle study. Microplastics and Nanoplastics 1: 1-13. Van Sickle, C. 1999. Southern Appalachian case study. Pages 472-488 in M.E. Jensen and P.S. Bourgeron, editors. A Guidebook for Integrated Ecological Assessment. Springer, New York, New York, USA. Wagner, S., P. Klöckner, B. Stier, M. Römer, B. Seiwert, T. Reemtsma, and C. Schmidt. 2019. Relationship between discharge and river plastic concentrations in a rural and an urban catchment. Environmental Science & Technology 53:10082-10091.

Methods

Identified MPs were recorded, quantified, and used for later data analysis relating discharge to MP concentration. A select number of MPs were sent for analysis using a Raman spectrometer to identify the specific chemical

> Fig. 1. Map of sample sites in the Chattooga watershed overlayed with National Land Cover Data. The watershed is primarily forested with scattered low density housing developments. The unincorporated town of Cashiers sits in the northeastern part of the watershed. QGIS.

Discussion and Conclusions

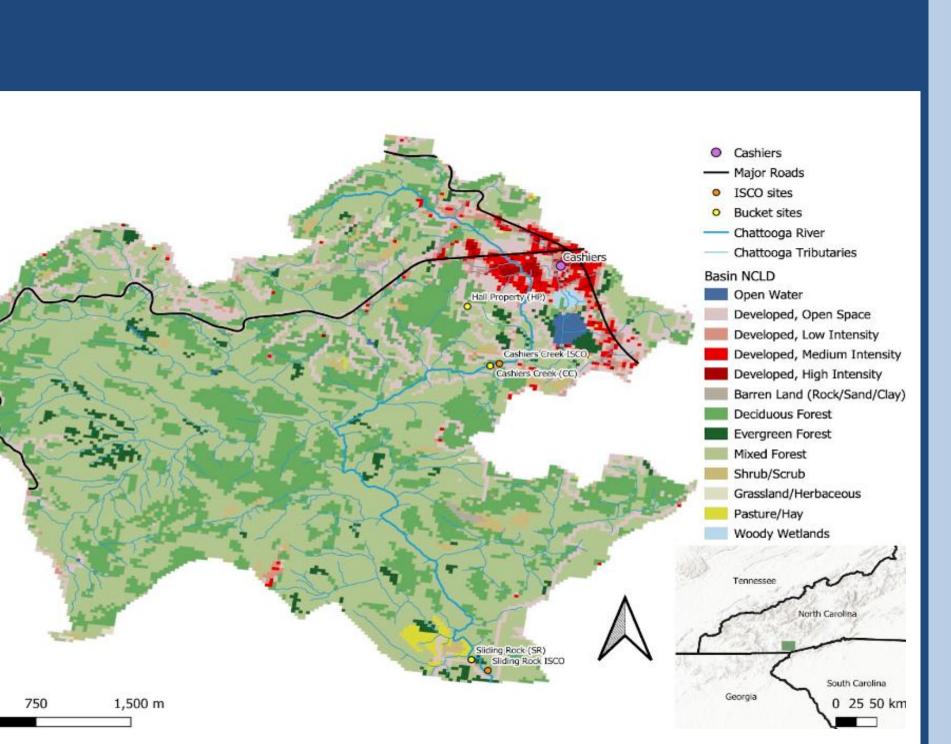
cubic meters per second) changed during the storm event

• No significant relationship between discharge and MP concentration was determined, however similarities between atmospheric and stream MPs indicate potential atmospheric circulation and

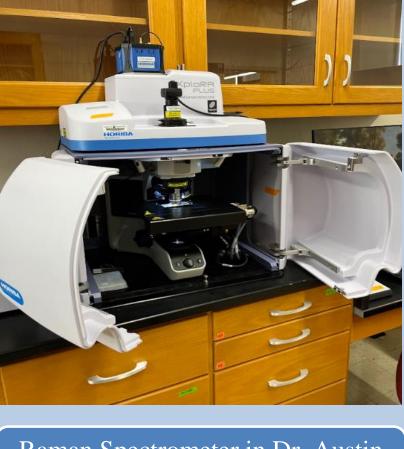
• Our study's estimated maximum number of MPs found in one sample was 5.08 × 104 MPs/m3 during storm flow. This result is magnitudes higher than other studies findings and indicates potential high contamination levels for the Chattooga river watershed. We confirmed approximately 82% of collected particles as anthropogenic microparticles using Raman spectroscopy. • MP contamination poses a unique threat to the biodiversity hotspot that is the southern Appalachians, the streams in this region are the "water towers" of the Southeast and should be monitored closely. • Significant MP contamination can occur during sampling and it is vital to correct for this contamination. This study is ongoing and additional work will help to delineate the sources, and drivers of MPs.

References









Raman Spectrometer in Dr. Austir Gray's lab at VT.

TABLE 1. Amount of atmospheric deposition (MPs/m²/day) at sites in the upper Chattooga River watershed, Jackson Co., NC between 1 and 21 October 2021. Sliding Rock and Hall Property sites were throughfall deposition. Deposition was highest at Cashiers Creek which was an open field site.

Location	Date Deployed	Date Collected	Total Time (days)	$\text{MPs}\;(m^2/\text{day})$
Sliding Rock	1 Oct	8 Oct	7.0	175.7
	8 Oct	15 Oct	6.9	1.1
	15 Oct	21 Oct	6.1	23.6
Cashiers Creek	1 Oct	8 Oct	6.9	190.8
	8 Oct	15 Oct	6.9	12.0
	15 Oct	21 Oct	6.1	8.7
Hall Property	1 Oct.	8 Oct	7.1	162.0
	8 Oct	15 Oct	6.9	12.0
	15 Oct	21 Oct	6.2	18.38