

# Using a TCR-transgenic CD4 T cell Approach to Guide Rational Chlamydia Vaccine Design



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Matthew Lu<sup>1</sup>, Taylor B. Poston<sup>2</sup>, Jenna Girardi<sup>2</sup>, Grace Polson<sup>3</sup>, & Toni Darville<sup>2</sup>

<sup>1</sup> Department of Biology, University of North Carolina at Chapel Hill, Chapel Hill, NC

<sup>2</sup> Department of Pediatrics, University of North Carolina at Chapel Hill, Chapel Hill, NC

<sup>3</sup> Department of Microbiology and Immunology, University of North Carolina at Chapel Hill, Chapel Hill, NC

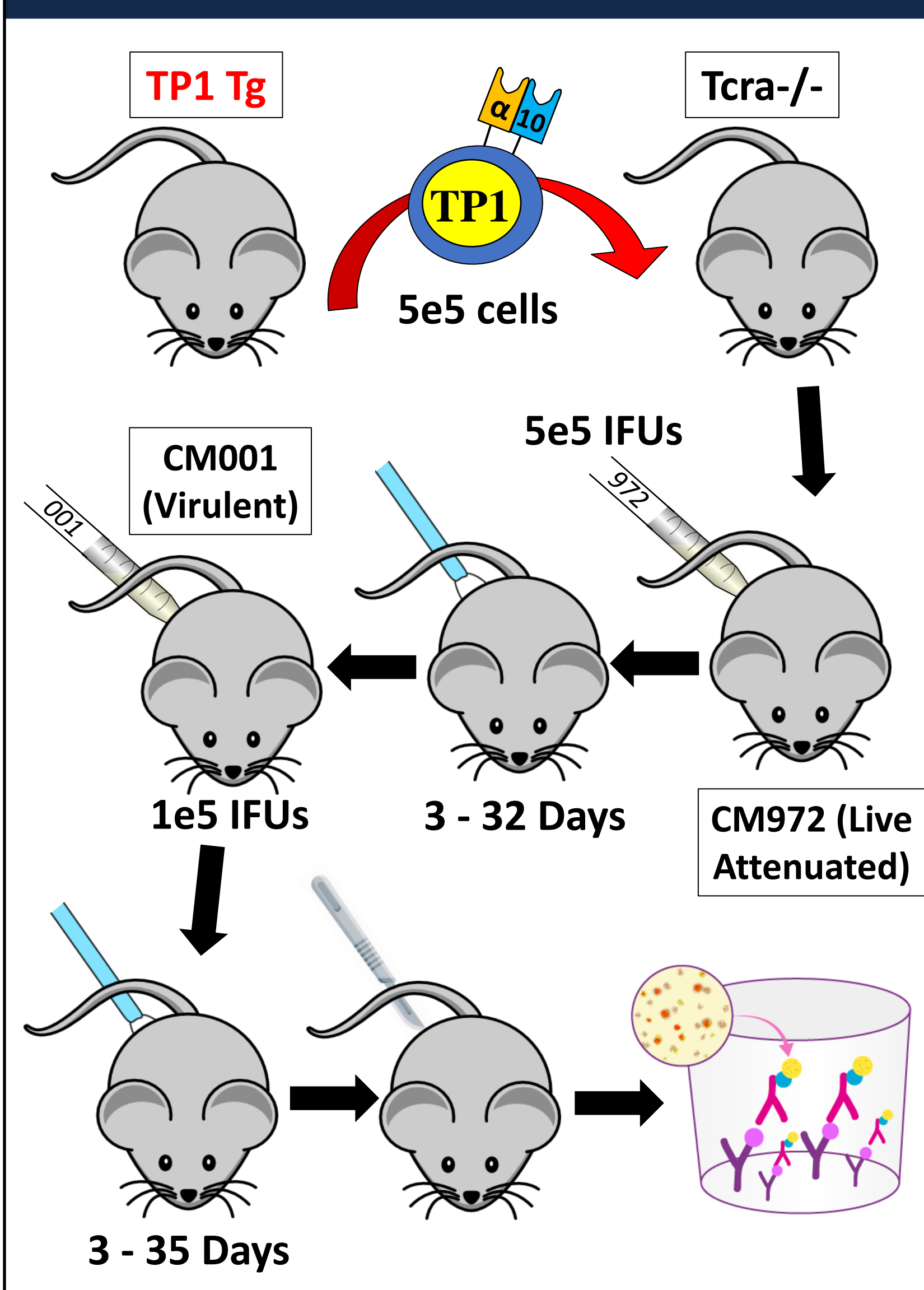


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## INTRODUCTION

- *Chlamydia trachomatis* is the most prevalent sexually transmitted bacterial infection globally.
- In 2018, there were 129 million new cases worldwide<sup>1</sup>.
- 70% of women and 50% of men are asymptomatic, which can lead to late or no treatment<sup>2</sup>.
- Untreated chlamydia can lead to serious symptoms such as pelvic inflammatory disease and infertility<sup>3</sup>.
- Previous research demonstrates that CD4+ T cells are critical for protection against infection.
- We developed a novel TCR transgenic mouse (TP1) model to investigate the protective capability of a monoclonal CD4+ T cell response to genital infection<sup>4</sup>.
- To determine this, we immunized TP1 recipient mice with live-attenuated *Chlamydia muridarum* (CM972) and later intravaginally challenged CM972-immunized mice with a virulent strain (CM001)<sup>5</sup>.

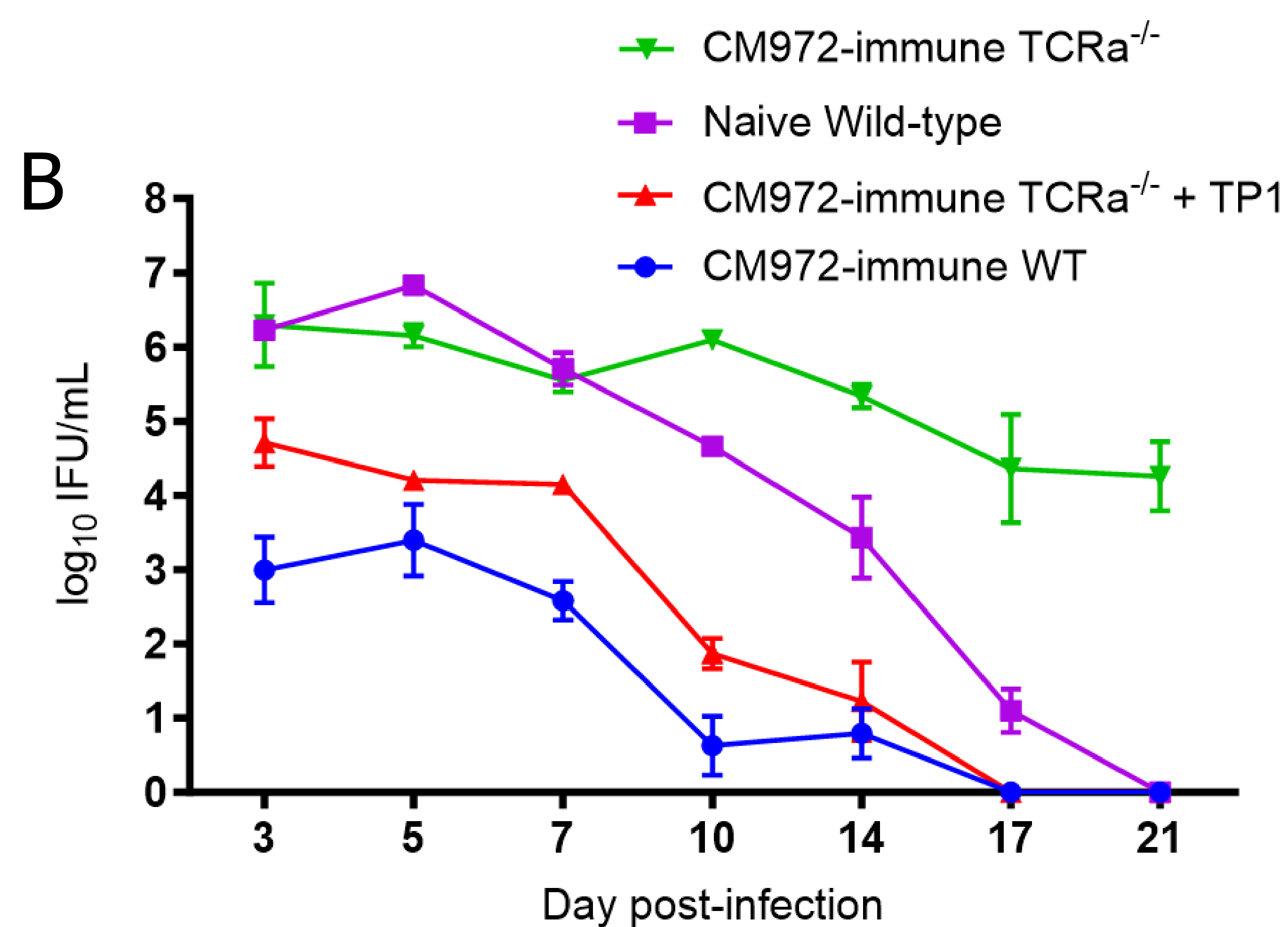
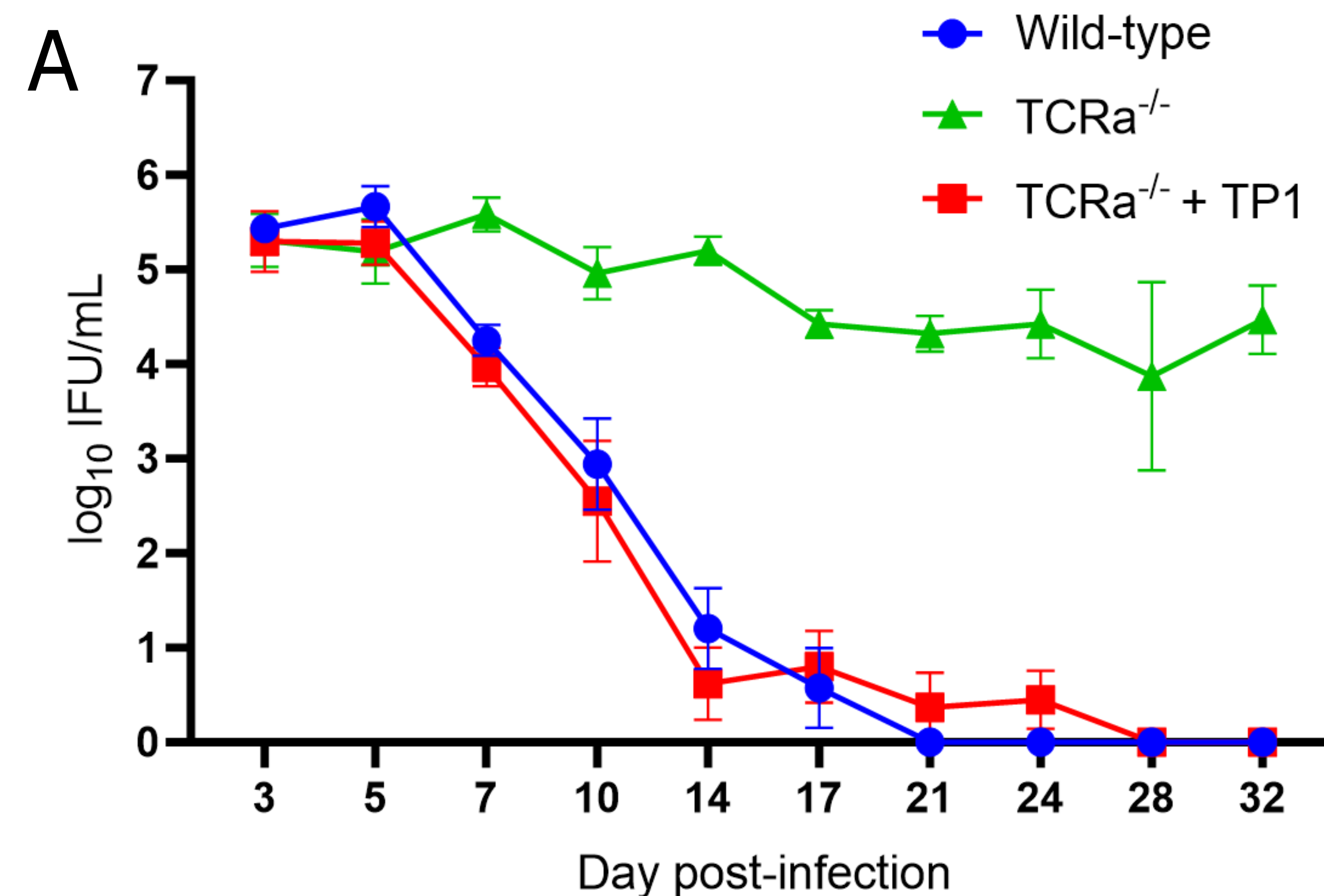
## METHODS



## HYPOTHESIS

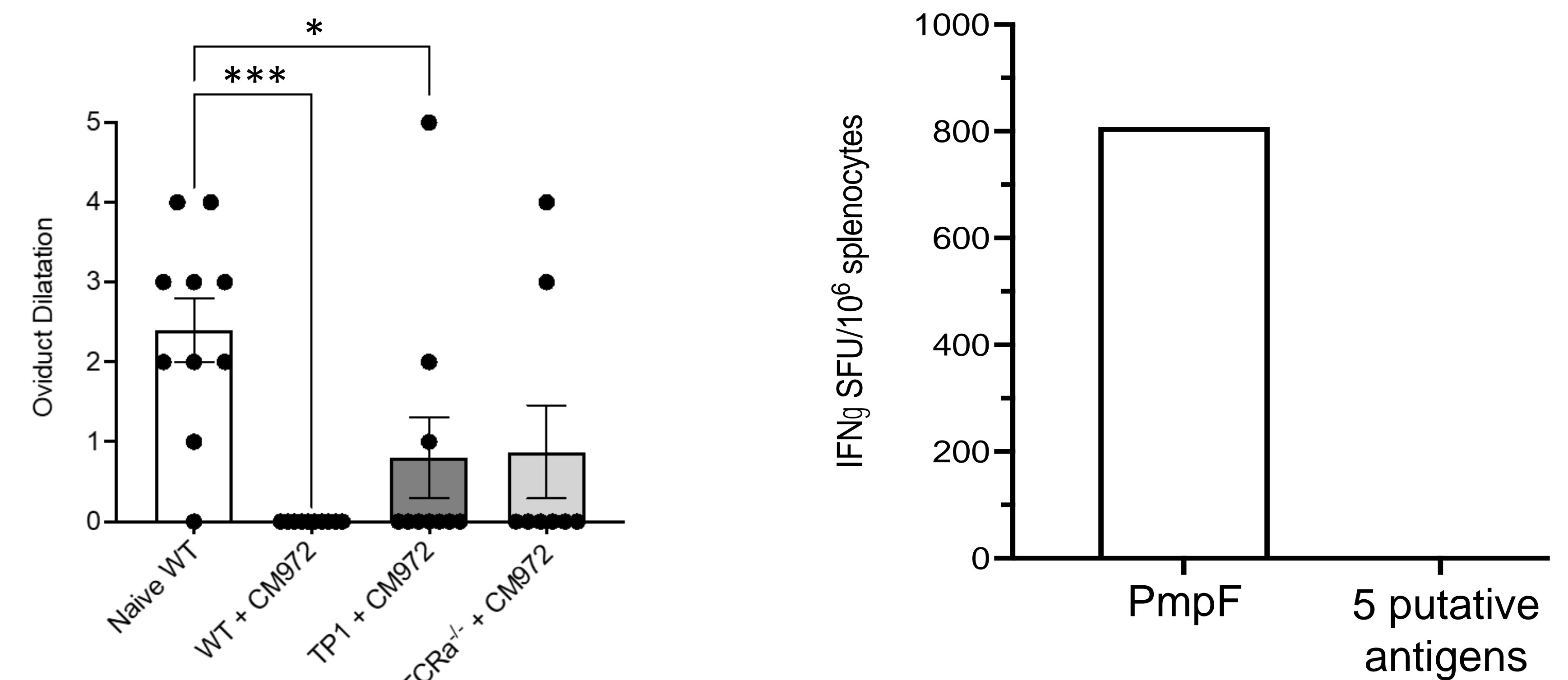
Memory TP1 cells specific for a single epitope will significantly reduce bacterial burden compared to naïve controls.

## RESULTS



**Fig. 1. Memory TP1 cells reduce chlamydial burden in genitally challenged mice.** Wild type (WT), TCRα<sup>-/-</sup>, and TCRα<sup>-/-</sup> with adoptively transferred TP1 cells were inoculated with *C. muridarum* strain CM972. (A) CM972 infection was monitored by IFU assay and mice were treated with doxycycline post-clearance. Mean ± SEM of 4-5 mice per group depicted. WT vs. TCRα<sup>-/-</sup> + TP1 ( $p=0.9280$ ), TCRα<sup>-/-</sup> + TP1 vs. TCRα<sup>-/-</sup> (-2.8 logs),  $p<0.0001$  by 2-way repeated measures ANOVA. (B) Naïve and CM972-immune mice were challenged with strain CM001. Infection was monitored as above. Immune WT vs. Immune TP1 (-0.82 log),  $p=0.0076$ ; Immunized TP1 vs. Naïve WT (-1.7 log),  $p<0.0001$ ; Immunized TP1 vs. Immunized TCRα<sup>-/-</sup> (-3.1 logs),  $p<0.0001$ ; Immunized WT vs. Naïve WT (-2.5 logs),  $p<0.0001$ , all by 2-way repeated measures ANOVA.

## RESULTS



**Fig 2. Memory TP1 cells reduce severity of oviduct pathology.** Mice from Fig 1B were sacrificed at 35 days post-secondary infection. Oviduct dilatation scores were determined by a pathologist blinded to the study design (n=8-10 oviducts per group). WT + CM972 vs. Naïve WT,  $p<0.0001$ ; TP1 + CM972 vs. Naïve WT,  $p<0.01$ , by Kruskal-Wallis test.

**Fig 3. TP1 cells are specific for polymorphic membrane protein F (PmpF).** Splenocytes from TCRα<sup>-/-</sup> mice that received TP1 cells followed by CM972 and CM001 infections were screened against a panel of Chlamydia antigens (MOMP, OmcB, PmpG, CPAF, Hsp60) by interferon-gamma ELISpot assay. SFU = spot-forming units.

## CONCLUSIONS & FUTURE DIRECTIONS

- Memory TP1 cells significantly reduced chlamydial burden compared to naïve wild-type T cells but were not as efficacious as memory wild-type T cells.
- Despite reducing burden by over 2-logs compared to naïve controls, memory TP1 cells did not prevent oviduct hydrosalpinx compared to immune controls.
- Antigen screening revealed that TP1 cells are specific for polymorphic membrane protein F (PmpF), an adhesin important for Chlamydia entry.
- We will next determine the specific PmpF peptide recognized by TP1 cells and perform preliminary PmpF vaccine studies.

## REFERENCES

- 1WHO. (2021). Global progress report on HIV, viral hepatitis and sexually transmitted infections, 2021 (p. 108). WHO. <https://www.who.int/publications/i/item/9789240027077>
- 2Laar, M. J. V. de, & Morré, S. A. (2007). Chlamydia: A major challenge for public health. *Eurosurveillance*, 12(10), 1-2. <https://doi.org/10.2807/esm.12.10.00735-en>
- 3Farris, C. M., & Morrison, R. P. (2011). Vaccination against Chlamydia Genital Infection Utilizing the Murine *C. muridarum* Model. *Infection and Immunity*, 79(3), 986-996. doi:10.1128/iai.00881-10
- 4A Chlamydia-Specific TCR-Transgenic Mouse Demonstrates Th1 Polyfunctionality with Enhanced Effector Function. *J Immunol*. 2017 Oct 15;199(8):2845-2854. doi: 10.4049/jimmunol.1700914.
- 5O'Connell, C. M., Ingalls, R. R., Andrews, C. W., Jr., Scurlock, A. M., & Darville, T. (2007). Plasmid-Deficient *Chlamydia muridarum* Fail to Induce Immune Pathology and Protect against Oviduct Disease1. *The Journal of Immunology*, 179(6), 4027-4034. <https://doi.org/10.4049/jimmunol.179.6.4027>

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