

## ENDOTHELIAL NUCLEAR PROTEINS SUN1 AND SUN2 REGULATE MICROTUBULE **ORGANIZATION IN ENDOTHELIAL CELLS**



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| Abstract   | Introduction   |   | Ν  | laterials an | d Methods   |  |  |  |
|--|--|---|--|--------------|---|--|--|--|
| Malfunctions in the LINC (Linker of Nucleoskeleton and Cytoskeleton) complex machinery lead to varying cases of muscular dystrophy, cardiac and skeletal muscle pathologies, and progeria. Progeria is a premature aging syndrome characterized by the development of atherosclerosis in young patients. Although endothelial cell (EC) dysfunction contributes to cardiovascular impairments in progeria patients, the role of the LINC complex in EC is still poorly understood. Our study examines how the depletion of LINC complex proteins affects microtubule organization in endothelial cells. Specifically, we investigate the unique and shared functions of SUN1, SUN2, and SYNE1 (nesprin-1) proteins. Through siRNA-mediated depletion, re-nucleation assays, and orbital flow experiments, the research identifies how SUN1, SUN2, and SYNE1 influence centrosome positioning and microtubule nucleation in cultured endothelial cells. Results indicate that the depletion of <i>SUN1</i> , <i>SUN2</i> , or <i>SYNE1</i> , individually or in combination, impacts centrosome distance from the nucleus, induces Golgi apparatus dispersal, and delays microtubule re-nucleation. The findings suggest that the LINC complex components are critical for the communication between the nucleus and the centrosome, which has implications in endothelial cell polarization and signal transduction, contributing to vascular biology and pathology. | <ul> <li>The LINC comp<br/>Cytoskeleton is on<br/>esprins located<br/>membrane, resp</li> <li>SUN2 is hypoth<br/>the behavior of<br/>forces between the<br/>such as sources</li> <li>Our lab showed<br/>dynamic and p<br/>preventing the<br/>(Nesprin-1) and<br/>such as SUN2.</li> <li>Preliminary resu<br/>SUN1, SUN2 is<br/>not as SUN2.</li> <li>Preliminary resu<br/>SUN1, SUN2 is<br/>not as SUN2.</li> <li>How do the LINC</li> </ul> | ults from our lab showed that unlike<br>regulates flow-induced endothelial<br>on, a process regulated by<br>anization in other cell types. <sup>3</sup><br>NC complex proteins SUN1, SUN2,<br>gulate microtubule organization and | Cell culture (HUVEC)<br>siRNA<br>Transfection<br>SiRNA<br>Transfection<br>Cell Splitting and<br>Seeding<br>Total Flow<br>Assay<br>Marenucleation<br>Marenucleation |              | Microtubule re-nucleation assay         Step 2       Step 3         Wash w/       PBS         Removes       At 37°C for time         Nocodazole       Culture media         PBS       Culture media         Vacation       Culture media         At 37°C for time       Culture         At 37°C for time |  |  |  |
| Results  |  |   |  |              |   |  |  |  |
| siRNA transfection successfully decreased SUN1, SUN2 and SYNE1 protein expression SUN1 and SUN2 regulate centrosome distance from the nucleus under flow   |  |   |  |              |   |  |  |  |

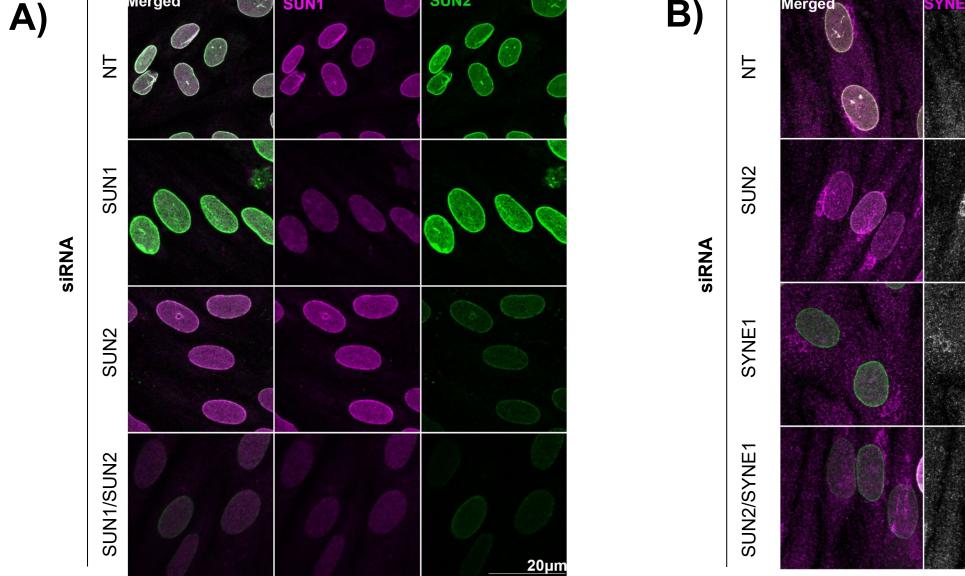
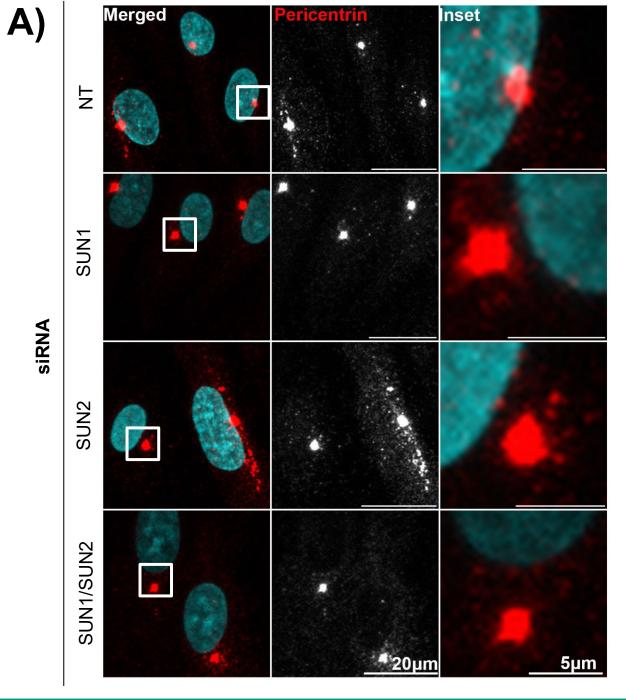


Figure 2. Analysis of SUN1, SUN2 and SYNE1 protein expression by immunostaining after treating Human Umbilical Vein Endothelial Cells (HUVEC) with *NT*, *SUN1*, *SUN2* or *SYNE1* siRNA and staining with SUN2 (in green), SUN1 (in pink) antibodies (A) or with SUN2 (in green) or SYNE1 (in pink) antibodies (B).



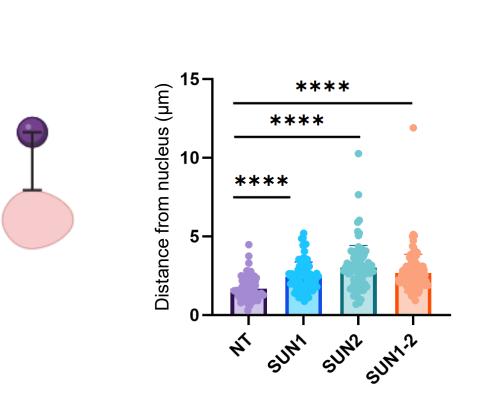


Figure 3. (A) HUVEC treated with NT, SUN1, and SUN2 siRNA were exposed to shear stress during 24hr using the orbital flow system, fixed with methanol and stained for pericentrin (in red, centrosome) and DAPI (in blue, nucleus). (B) Quantification of distance between the nucleus and the centrosome for all siRNA conditions showing increased distance between the nucleus and the centrosome after individual or combined depletion of SUN1 or SUN2. Unpaired two-tailed t-test. \*\*\*\*P<0.0001. Scale bars: 20 μm or 5 μm.

🔲 NT

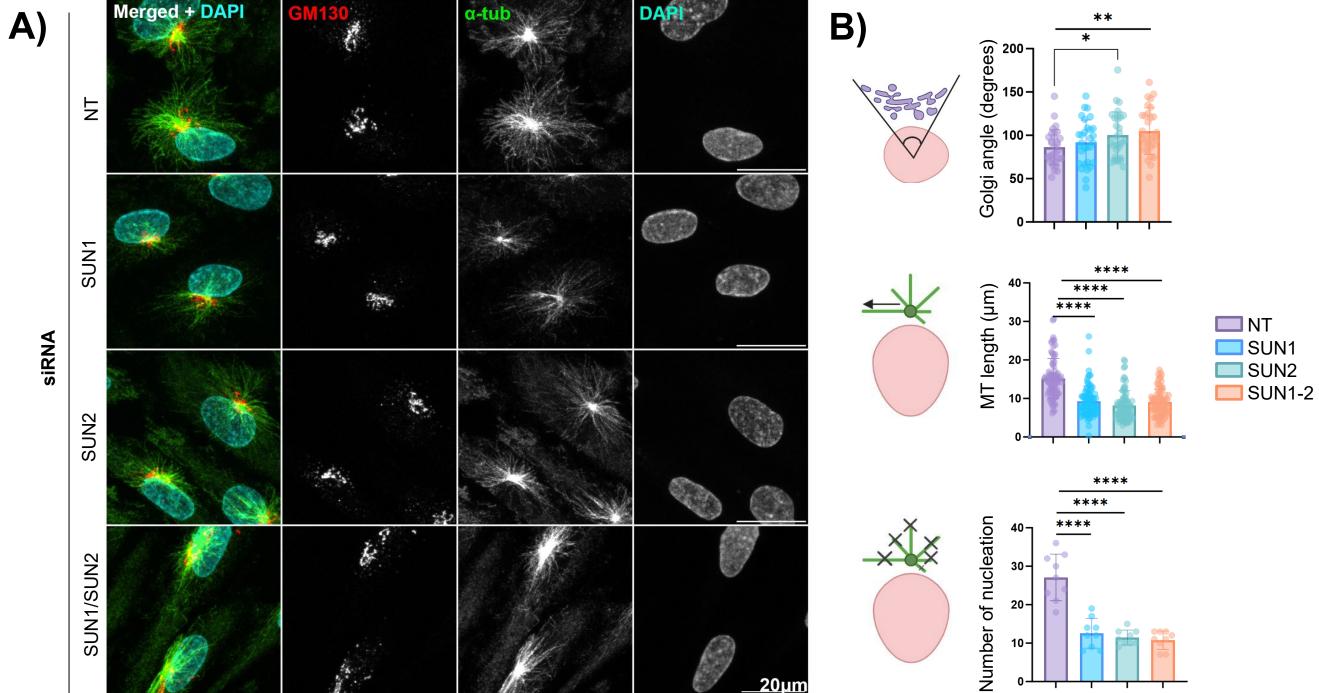
SYNE1

🔲 SUN2

SYNE1

SUN2

## SUN1 and SUN2 regulate microtubule dynamics during re-nucleation assay



## SUN2 and SYNE1 show synergistic effects on microtubule dynamics

B)

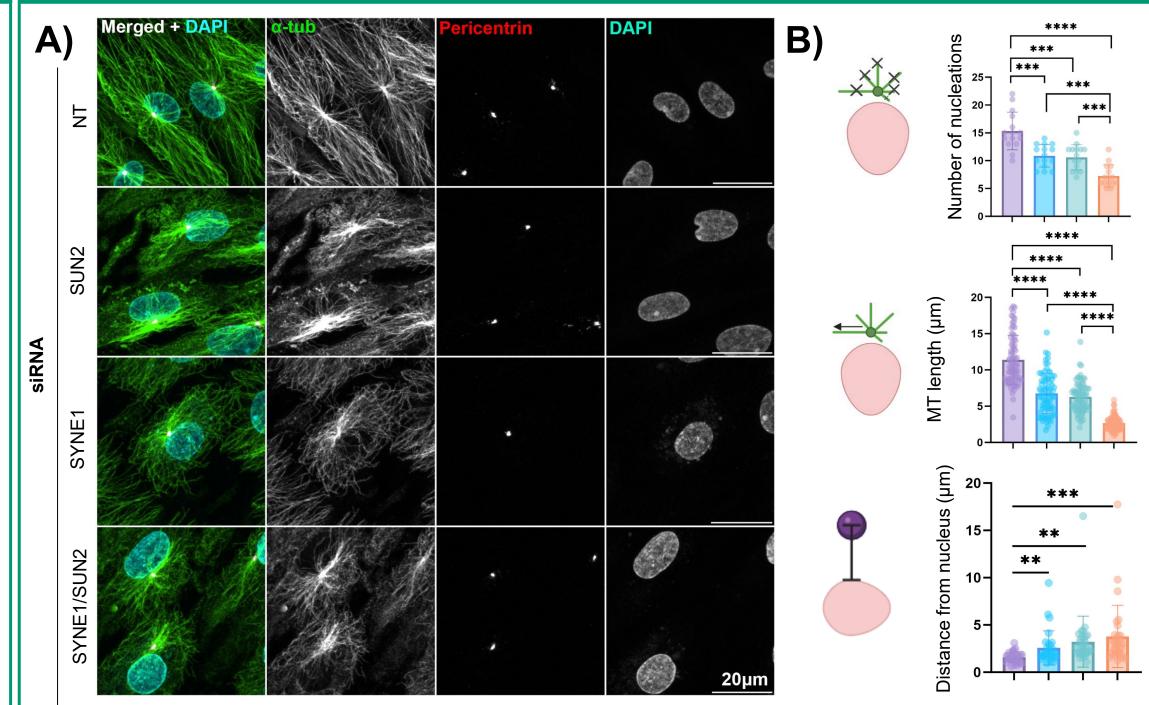


Figure 4. (A) HUVEC transfected with NT, SUN1 and SUN2 siRNA were allowed to recover for 5 min after Nocodazole treatment. Cells were then fixed and stained for α-tubulin (in green), GM130 (in red, Golgi apparatus) and DAPI (in blue, nucleus). (B) Quantification of Golgi apparatus angle, microtubule length and nucleation for all siRNA conditions showing increased Golgi dispersal in SUN2-depleted cells and delayed microtubule re-nucleation after individual depletion of SUN1 or SUN2. SUN1/SUN2 co-depletion showed a similar effect to the individual depletion of SUN2. Unpaired two-tailed t-test. \*P<0.05; \*\*P<0.01; \*\*\*P<0.001; \*\*\*\*P<0.001; Scale bars: 20 µm.

Figure 5. (A) HUVEC transfected with NT, SUN2 and SYNE1 siRNA were allowed to recover for 10 min after Nocodazole treatment. Cells were then fixed and stained for α-tubulin (in green), pericentrin (in red, centrosome) and DAPI (in blue, nucleus). (B) Quantification of number of renucleations, microtubule length, and distance between nucleus and centrosome for all siRNA conditions showing delayed microtubule re-nucleation and increased distance between the nucleus and the centrosome after individual depletion of SUN2 or SYNE1. SUN2/SYNE1 co-depletion showed additive effect. Unpaired two*tailed t-test.* \*\*P<0.01; \*\*\*P<0.001; \*\*\*\*P<0.0001; Scale bars: 20 μm.

| Conclusions | <b>Future Directions</b>   | References  |  |
|-------------|--|---|--|
|             | <ul> <li>Further investigation of the role of SYNE1 in nucleocytoplasmic communication, specifically in interactions with the Golgi Apparatus.</li> <li>Exploring the role of SUN1/2 in nucleocytoplasmic communication via non-centrosomal microtubules</li> <li>Investigation of motor proteins adaptors and their relationship to LINC Complex</li> <li>Investigation of impacts of SUN2/SYNE1 on actin cytoskeleton</li> <li>Mank you to Dr. Victoria Bautch and Dr. Pauline Bougaran for their invaluable mentorship throughout the research project.</li> <li>Funding source: NIH-NHLBI R35-HL139950-01</li> </ul> | <ol> <li>King, M. C. Dynamic Regulation of LINC<br/>Complex Composition and Function<br/>across Tissues and Contexts. FEBS<br/>Letters 2023, 597 (22), 2823–2832.<br/><u>https://doi.org/10.1002/1873-<br/>3468.14757</u>.</li> <li>Buglak, D. B., Kulikauskas, M. R., Liu,<br/>Z., Gold, A. L., Marvin, A. P., Burciu, A.,<br/>Tanke, N. T., Ricketts, S. N., Kinghorn,<br/>K., Oatley, M., Johnson, B. N.,<br/>Bougaran, P., Shiau, C. E., Rogers, S.<br/>L., &amp; amp; <b>Bautch, V. L.</b> (2021). Nuclear<br/>SUN1 stabilizes endothelial cell junctions<br/>via microtubules to regulate blood vessel<br/>formation.<br/><u>https://doi.org/10.1101/2021.08.11.45598</u><br/><u>0</u></li> <li>Martin, M.; Veloso, A.; Wu, J.; Katrukha,<br/>E. A.; Akhmanova, A. Control of<br/>Endothelial Cell Polarity and Sprouting<br/>Angiogenesis by Non-Centrosomal<br/>Microtubules. eLife 2018, 7, e33864.<br/><u>https://doi.org/10.7554/eLife.33864.</u></li> </ol> |  |